



**Blood Bank, We Have a  
Problem: Working together to  
Optimize Antibody Resolution**

Nancy Mullis, MT(ASCP)SBB

LeeAnn Prihoda, MEd, MT(ASCP)SBB

American Red Cross, Southern Region

Immunohematology Reference Laboratory



“Is there anything I can do here?”

## **Phone consultation**

- Supervisor/Lead Tech
- If patient was previously evaluated by ARC, ARC# or date tested (if recent)
- Fax worksheets for review
- Based on serologic problem, you may be able to avoid sending sample to Ref Lab

Recognizing the problem is  
always the first step....

## Evaluating the problem

- Patients with previous evaluations
  - Review report or call Ref Lab for consultation
    - What was identified? Has reactivity changed?
    - What test method was used for identification and for ruling out other antibodies?
    - What resources do you have for testing?
    - Do you need antigen-negative units?

# Evaluating the problem

- Patients with no previous evaluations
  - Obtain thorough patient history
  - What antibodies can be ruled out? Are there any obvious specificities? If crossmatches were done, what percent are compatible?
- Is there a way to safely provide transfusion pending results of antibody ID?
  - Have common alloantibodies been excluded?
  - Are there compatible crossmatches?

# Other resources I would highly recommend



# “Unexplained Reactivity”

## Reactions with few cells on the panel

- Strong reactions most likely due to antibody to a LIA
- Weaker reactions
  - Antibodies to white cell antigens (Bg)
  - Antibody reacting only with cells that have stronger expression of antigen (I, IH, P<sub>1</sub>, D or other Rh specificities)
  - Antibodies showing dosage (-M, -Jk<sup>a</sup>)
  - Developing antibodies, ???

# What to do...

- Are you able to rule out the presence of common alloantibodies?
  - Use cells with double dose expression
    - r/o of anti-K an exception. K+k+ cells are acceptable.
- Do you have compatible crossmatches?
  - Crossmatch/screen using method by which the reactivity was detected

If **Yes to both**, it is acceptable to transfuse the patient....investigation of the reactivity may continue (get adequate samples first!)

# The Low-Down...

D	C	E	c	e	K	S	s	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	Gel
+	0	0	+	+	0	0	+	0	+	0	+	0
+	0	+	+	0	0	0	+	0	+	0	+	3+
0	0	0	+	+	+	0	+	+	0	+	0	0
0	0	0	+	+	0	+	0	0	+	0	+	0
+	0	+	+	0	0	0	+	0	+	+	0	0
+	+	0	0	+	0	+	+	0	+	0	+	0
0	0	+	+	+	0	0	+	0	+	0	+	3+



# The Low-Down...

D	C	E	c	e	K	S	s	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	Gel
+	+	0	0	+	0	0	+	0	+	0	+	0
+	0	+	+	0	0	0	+	0	+	0	+	3+
0	0	0	+	+	+	0	+	+	0	+	0	0
0	0	0	+	+	0	+	0	0	+	0	+	0
+	0	+	+	0	0	0	+	0	+	+	0	0
+	+	0	0	+	+	+	0	+	0	0	+	0
0	0	+	+	+	0	0	+	0	+	0	+	3+

# The Low-Down...on lows

DONOR	(Bg <sup>a</sup> , Bg <sup>b</sup> , Bg <sup>c</sup> )	Coa	Cob	Sda	Ytb
101984	0		0		
107952	0		0		
108226	0		0		
111208	0				
111209	0				
111247	0				
111302	0			0	
111329	0		+		
111375	0		0		
111444	0		0		
111931	0				
112858	0		0		

# Common LIA on your panels

- Rh: V, Go<sup>a</sup>, C<sup>w</sup>
- Kell: Kp<sup>a</sup>, Js<sup>a</sup>
- Lu<sup>a</sup>
- Yb<sup>b</sup>
- Co<sup>b</sup>
- He
- Dj<sup>a</sup>
- And hundreds more that aren't listed!

# Case of Mistaken Identity

- 15 y/o male with sickle cell disease
- Admitted with “sickle cell crisis”
- ABO/Rh: A pos, rec'd only C-E-K- units per PFL protocol
- Last transfusion in Florida, 10 days before sample submitted for Reference evaluation
- Per hospital in Florida, anti-E and –Jk<sup>a</sup> were identified at a Florida Ref Lab. Patient rec'd C-E-K-Jk(a-) red cells.

# D.C.

- Initial results:
  - DAT negative (weak pos DAT reported by Florida hospital)
  - Anti-Jk<sup>a</sup> detected in PEG. Anti-E was not detected in PEG.
  - Gel panel subsequently tested to try and detect anti-E reported by Florida Reference Lab.

D	C	E	c	e	K	S	s	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	Gel
+	0	0	+	+	0	0	+	0	+	0	+	2+
+	0	+	+	0	0	0	+	0	+	0	+	3+
0	0	0	+	+	+	0	+	+	0	0	+	0
0	0	0	+	+	0	+	0	0	+	0	+	0
+	0	+	+	0	0	0	+	0	+	0	+	3+
+	+	0	0	+	0	+	+	0	+	0	+	2+
0	0	+	+	+	0	0	+	0	+	0	+	0

D	C	E	c	e	K	S	s	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	Gel
+	0	0	+	+	0	0	+	0	+	0	+	2+
+	0	+	+	0	0	0	+	0	+	0	+	3+
0	0	0	+	+	+	0	+	+	0	0	+	0
0	0	0	+	+	0	+	0	0	+	0	+	0
+	0	+	+	0	0	0	+	0	+	0	+	3+
+	+	0	0	+	0	+	+	0	+	0	+	2+
0	0	+	+	+	0	0	+	0	+	0	+	0

D	C	E	c	e	K	S	s	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	Gel
+	0	0	+	+	0	0	+	0	+	0	+	2+
+	0	+	+	0	0	0	+	0	+	0	+	3+
0	0	0	+	+	+	0	+	+	0	0	+	0
0	0	0	+	+	0	+	0	0	+	0	+	0
+	0	+	+	0	0	0	+	0	+	0	+	3+
+	+	0	0	+	0	+	+	0	+	0	+	2+
0	0	+	+	+	0	0	+	0	+	0	+	0



# Estimated number of available antigen sites on D+ red cells

Issitt PD, Anstee DJ. Applied Blood Group Serology, 4<sup>th</sup> edition

Phenotype		# of D antigen sites
cDE/cDE	$R_2R_2$	16000-33500
CDe/cDE	$R_1R_2$	23000-31000
CDe/CDe	$R_1R_1$	14500-19500
cDE/cde	$R_2r$	14000-16500
cDe/cde	$R_0r$	12000-20000

# D.C.

- Antibody ID (plasma): anti-D and  $-Jk^a$
- Eluate prepared (even though tube DAT was negative), demonstrated anti-D
  - Auto control in Gel was 2+
- Patient is D+
  - Auto vs. Alloimmune anti-D?
  - Eluate performed on patient cells obtained after cell separation was non-reactive, suggestive of alloimmune anti-D.

## D.C.

- Samples sent for molecular testing to determine if patient has variant D gene.
- Molecular lab determined patient inherited a partial D antigen, and anti-D was determined to be alloimmune.
- When identified in Florida, anti-D was likely weaker and only reactive with R<sub>2</sub>R<sub>2</sub> cells (thus appeared to be anti-E)
- **Patient may have been having a delayed transfusion reaction due to anti-D**

# More Unexplained Reactions

- Reactions with most or all panel cells
  - Antibody to HIA
  - Warm or Cold-reactive autoantibody
  - Multiple alloantibodies
  - Extraneous reactivity (antibody to component of test system: preservatives, diluent solutions, antibiotics, enhancement, etc.)

# Recognizing Media-dependent reactivity

- Test sample using other methods, similar in sensitivity and reaction phase
  - LISS dependent antibodies – test @ 37C phase using albumin
  - Gel dependent antibodies
    - test using PEG or other enhancement media, at antiglobulin-phase
    - Test pre-diluted Ortho cells vs 0.8% cells prepared in lab from another source (donors, other screening cells or panel cells)

# Much Ado about Nothing Case of Missing Reactions

	IS	PEG IAT
SI	0	0
SII	0	0
SIII	0	0
AC	0	0

	LISS 37	LISS IAT
SI	0	0
SII	0	0
SIII	0	0
AC	0	0

# 0.8% Panel

D	C	E	c	e	K	S	s	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	Gel
+	0	0	+	+	0	0	+	0	+	0	+	3+
+	0	+	+	0	0	0	+	0	+	0	+	3+
0	0	0	+	+	+	0	+	+	0	+	0	3+
0	0	0	+	+	0	+	0	0	+	0	+	3+
+	0	+	+	0	0	0	+	0	+	+	0	3+
+	+	0	0	+	0	+	+	0	+	0	+	3+
Auto Control												0

# Other reagent cells diluted to 0.8%

	<b>Phenotype</b>	<b>Gel</b>
1	<b>Phen. Sim Reagent cell</b>	<b>0</b>
2	<b>Phen. Sim. Donor unit</b>	<b>0</b>
3	<b>Kp(b-)</b>	<b>0</b>
4	<b>Lu(a-b-)</b>	<b>0</b>
5	<b>Knops null</b>	<b>0</b>

- Reactions only noted with pre-diluted cells from manufacturer
- Antibody most likely directed at preservative or other component of pre-diluted panel cells



# Reading the ARC final report

**Serum Antibody:** Warm reactive autoantibody

**Reactive By:** LISS 37C/antiglobulin; PEG/antiglobulin

- **Additional Comments:** The patient's serum was adsorbed x2 at 37C with selected allogeneic red cells following papain treatment. The adsorbed serum was then tested with reagent red cells and was non-reactive. The presence of antibodies to common red cell antigens was excluded by PEG/IAT. However, antibodies to high incidence antigens may not be detected when testing allogeneic-adsorbed serum.

# Reading the ARC Final Report

**Serum Antibody:** Warm reactive autoantibody

**Reactive By:** PEG/antiglobulin, Gel test (IgG)

The presence of alloantibodies to common red cell antigens was excluded by LISS/IAT.

# Reading the ARC Final Report

**Serum Antibody:** Anti-Fy<sup>a</sup>

**Reactive By:** PEG/antiglobulin

Previously identified anti-C, -E, -S, and -Jk<sup>b</sup> were not reconfirmed at this time. The presence of alloantibodies to common red cell antigens was excluded by PEG/IAT.

# What to consider...

- What needs to be ruled out?
  - Patient has anti-C, -E, -S, -Fy<sup>a</sup>, Jk<sup>b</sup> already
- Patient's phenotype:  
C-E-c+e+; K-; Fy(a-b+); Jk(a+b-); S-s+

# What to consider...

- What needs to be ruled out?
  - Patient has anti-C, -E, -S, -Fy<sup>a</sup>, Jk<sup>b</sup> already
- Patient's phenotype:  
**C-E-c+e+; K-; Fy(a-b+); Jk(a+b-); S-s+**
- Anti-K needs to be ruled out.....or does it?
- Other potential problems to be considered
  - Autoantibodies, “HTLA” antibodies, lows, etc.
  - If you don't have a negative cell on the screen, test one or more phenotypically similar cells from a panel. If negative, it is probably “safe” to order phenotyped units w/o sending sample

# Submitting Samples to a Reference Laboratory

## What is important to know??

- Patient history
  - Previously identified antibodies
  - Transfusion history, esp. date of last trx
  - Diagnosis
  - Has the patient been seen at another hospital? – when, where
  - Pregnancy history
  - Medications / IV therapy

# Look away from the computer

## **Other resources for information:**

- Ask the patient or the patient's family
  - “have you ever been transfused?”
  - “have you had any problems being transfused”
  - “where have you been transfused?”
- Physician's admitting H&P may be on patient's chart - lots of info here!
- Nursing may be able to assist

# Other important information...

- Patient demographics (sex, age, race)
- What is the serologic problem you are seeing?
  - Send copies of worksheets (panels, ABSC)
  - Describe testing you have done (method(s) used, strength of reactivity, estimated prevalence of reactivity, phase(s) of reactivity, etc.)



# Things that bring work to a halt

- Mislabeled samples/ discrepant requests
  - Patient name must match exactly
  - ID# must match exactly
  - Collection date must be on sample, must match date on request
- Insufficient sample
  - Slows work down
  - Transit time for additional samples adds to overall turn-around-time

# More Speed Bumps....

- Missing information
- Inaccurate information
  - Especially transfusion history (basis for cell separations, eluates, interpretation of screen...)
- Multiple phone calls
  - Call us if you have more information or the patient's status has changed
- Changing the order multiple times
  - If you're unsure about how many units are needed, or if special requirements (CMV, IRR, AG-) are needed, PLEASE get this worked out before placing the order!

# New ARC Request Form!!

American Red Cross  
Biomedical Services  
Washington, DC 20006

## Immunoematology Consultation Request

Southern Region

CLIA # 11-D0699333

Reference Lab contact numbers: Phone: (404) 253-5580 or 1-800-884-2710 ext. 5580 Fax: (404) 876-6984

See page 2 for instructions, sample types and tube labeling requirements

### 1 Call Reference Laboratory before sending sample

Reference Lab person contacted: \_\_\_\_\_ Date/Time contacted: \_\_\_\_\_

### 2 Submitting Facility Information

Facility Name/ID: \_\_\_\_\_ Request Date: \_\_\_\_\_  
Facility Address: \_\_\_\_\_ City/State: \_\_\_\_\_ Zip: \_\_\_\_\_  
Blood Bank Contact: \_\_\_\_\_  
Blood Bank Phone #: \_\_\_\_\_ Requesting Physician: \_\_\_\_\_  
Blood Bank Fax #: \_\_\_\_\_

### 3 Patient Information

Patient Name: \_\_\_\_\_ Patient ID: \_\_\_\_\_  
Birth Date/Age: \_\_\_\_\_ Race: \_\_\_\_\_ Gender: M  F   
Specimen Date: \_\_\_\_\_ ABO/Rh: \_\_\_\_\_  
Diagnosis: \_\_\_\_\_ Hgb/Hct: \_\_\_\_\_  
Medications: \_\_\_\_\_  
Additional information: \_\_\_\_\_

Transfusion History: No record

Within last 3 months: No  Yes  ► Dates / products: \_\_\_\_\_

Prior to last 3 months: No  Yes  ► Dates / products: \_\_\_\_\_

Pregnancy History: Number: \_\_\_\_\_ Currently pregnant? No  Yes  ► Due date: \_\_\_\_\_

Known RBC antibody(ies) Anti -D  -C  -c  -E  -e  -K  -Fy<sup>a</sup>  -Fy<sup>b</sup>  -Jk<sup>a</sup>  -Jk<sup>b</sup>  -S  -s

Other (list): \_\_\_\_\_

**4 Test Request** Note: STAT and/or after-hours charges may apply. Date/time needed: \_\_\_\_\_

**Routine**  Non-urgent, stable situations (examples include routine dialysis or elective surgery).

**ASAP**  Non-emergent situations where a shortened turn around time would be beneficial (example: low hemoglobin).

**STAT**  Status of the patient is extremely unstable. This is an urgent request (example: actively bleeding patient).

**Investigation Requested:** (Check all that apply)

ABO/Rh typing <input type="checkbox"/>	Incompatible crossmatch <input type="checkbox"/>	ABO/Rh typing Discrepancy <input type="checkbox"/>
Positive DAT <input type="checkbox"/>	Suspected transfusion reaction <input type="checkbox"/>	Partners for Life Evaluation <input type="checkbox"/>
Antibody identification <input type="checkbox"/>	Hemolytic Disease of the Fetus & Newborn <input type="checkbox"/>	<b>OTHER SPECIFY:</b>

**Products Requested for this patient:** (Check all that apply)

**Product Attributes:** (Check all that apply)

<b>CROSSMATCHED RBCs</b> # units: _____ Date/Time needed: _____ OTHER (SPECIFY): _____	<b>ANTIGEN-NEGATIVE RBCs</b> # units: _____ Date/Time needed: _____ Negative for: (Circle) C c E e K Fy <sup>a</sup> Fy <sup>b</sup> Jk <sup>a</sup> Jk <sup>b</sup> S s Other: _____	CMV-negative <input type="checkbox"/> Leukoreduced <input type="checkbox"/> Irradiated <input type="checkbox"/> Hemoglobin S-negative RBC <input type="checkbox"/> OTHER SPECIFY: _____
--	---	---

**5 Summary of Antibody Testing Results:** Please provide copies of blood bank test results.

Tube: LISS <input type="checkbox"/> PEG <input type="checkbox"/> ENZ <input type="checkbox"/> IS 37C AHG Other: <input type="checkbox"/> _____ Gel <input type="checkbox"/> Solid Phase <input type="checkbox"/>	I _____ II _____ III _____ AHG Used: Polyspecific <input type="checkbox"/> IgG <input type="checkbox"/>	<b>CROSSMATCH RESULT</b> # Compatible Donors _____ # Incompatible Donors _____ DAT: _____
---	--	--

American Red Cross  
Biomedical Services  
Washington, DC 20006

## Immunochemistry Consultation Request

Southern Region

CLIA # 11-D0699333

### Instructions for submitting samples for a Reference Laboratory investigation:

1. Obtain an appropriate sample and label it according to the instructions in the table below.
2. Prepare the sample for shipping according to the instructions in the table below.

#### Sample and Tube Label Requirements

##### Sample Type:

20 mL EDTA or combination of clotted and EDTA

- Serum separator tubes are **NOT** acceptable
- If recently transfused, include pre-transfusion red cell sample, if available

##### Label Requirements:

Sample Label **MUST** include:

- Patient's First and Last Names
- Patient's ID Number
- Date collected

No special preparation of the patient is needed prior to collecting samples.

**NOTE: IMPROPERLY LABELED SAMPLES WILL NOT BE TESTED!!**

##### Shipping Instructions:

Contact the Reference Lab if assistance is needed for sample transportation.

3. Complete the form on the reverse side.
  4. Contact the reference lab at the phone number on the top of the form prior to sending the sample.
  5. Record the name of the person contacted and the date and time of the notification.
  6. Follow any additional instructions noted in the ARC Facility-specific area below.
  7. Submit the sample and a completed Consultation Request to the ARC.
-

### Segment Compatibility Test Request/Report

When submitting segments for testing, please verify the following:

1. If possible, two segments have been submitted for each unit.
2. Each segment submitted has a complete segment number.
3. Each segment is labeled with the corresponding whole blood number.
4. The first three columns in the table below have been completed.

**Patient Name:** \_\_\_\_\_

**Patient ID #** \_\_\_\_\_

*A printed label may be used to provide this information.*

**NOTE: If segments are from units tagged antigen negative by American Red Cross, please include a photocopy of units with tag to prevent duplicate testing and charges.**

**NOTE: shaded area for ARC use only**

Unit number	Segment number	ABO & Rh	Crossmatch Result:	Clerical Review	Date
			<input type="checkbox"/> Compatible with <input type="checkbox"/> Least Incompatible with <input type="checkbox"/> Incompatible with		
1			<input type="checkbox"/> neat serum <input type="checkbox"/> allo absorbed <input type="checkbox"/> auto absorbed	/	
2			<input type="checkbox"/> neat serum <input type="checkbox"/> allo absorbed <input type="checkbox"/> auto absorbed	/	
3			<input type="checkbox"/> neat serum <input type="checkbox"/> allo absorbed <input type="checkbox"/> auto absorbed	/	
4			<input type="checkbox"/> neat serum <input type="checkbox"/> allo absorbed <input type="checkbox"/> auto absorbed	/	
5			<input type="checkbox"/> neat serum <input type="checkbox"/> allo absorbed <input type="checkbox"/> auto absorbed	/	
6			<input type="checkbox"/> neat serum <input type="checkbox"/> allo absorbed <input type="checkbox"/> auto absorbed	/	

Next, the fun stuff!



# Case J.G.

- Patient J.G.
  - African American male, DOB 12/7/53
  - Dx: Multi-organ failure, Hemolytic process
  - No transfusions in last 3 months
  - Hgb 7.9; submitted for antibody identification and 2 units crossmatched red cells



# J.G. Initial Results

Anti-A	Anti-B	Anti-D	Rh Ctl	A <sub>1</sub> cells	B cells
4+	0	4+	0	3+	4+

**DAT**

PS	-IgG	-C3	Control
1+	1+	0	0

# J.G. Initial Results

	<b>IS</b>	<b>PEG IAT</b>
<b>SI</b>	<b>2+s</b>	<b>3+</b>
<b>SII</b>	<b>2+s</b>	<b>3+</b>
<b>SIII</b>	<b>0</b>	<b>0</b>
<b>AC</b>	<b>0</b>	<b>1+</b>

	<b>LISS 37</b>	<b>LISS IAT</b>
<b>SI</b>	<b>3+</b>	<b>4+</b>
<b>SII</b>	<b>3+</b>	<b>3+</b>
<b>SIII</b>	<b>0</b>	<b>0</b>
<b>AC</b>	<b>0</b>	<b>1+</b>

# To Summarize

- ABO discrepancy
- Positive DAT
- Positive Antibody Screen
  - Phenotype patient cells
  - Identify direct agglutinating/IAT antibody
  - Prepare and test eluate

J.G.

**Phenotype:**

**C-E-c+e+; K-; Fy(a-b-); Jk(a+b-);**

**S-s+; M-; Le(a-b-); A<sub>1</sub>-**

J.G.

	Rh	K	M	N	Le <sup>a</sup>	Le <sup>b</sup>	P <sub>1</sub>
SI	R1R1	+	+	0	0	+	0
SII	R2R2	0	+	+	+	0	+
SIII	rr	0	0	+	0	+	+

J.G.

	Rh	K	M	N	Le <sup>a</sup>	Le <sup>b</sup>	P <sub>1</sub>
SI	R1R1	+	+	0	0	+	0
SII	R2R2	0	+	+	+	0	+
SIII	rr	0	0	+	0	+	+

# Panel Results - Summary

	<b>Phenotype</b>	<b>IS</b>	<b>LISS 37</b>	<b>LISS IAT</b>
1	<b>R1R1 M-</b>	<b>0</b>	<b>0</b>	<b>0</b>
2	<b>R2R2 M-</b>	<b>0</b>	<b>0</b>	<b>0</b>
3	<b>I- M-</b>	<b>0</b>	<b>0</b>	<b>0</b>
4	<b>rr M+</b>	<b>1+s</b>	<b>2+</b>	<b>2+</b>

# Panel Results - Summary

	Phenotype	IS	LISS 37	LISS IAT
1	R1R1 M-	0	0	0
2	R2R2 M-	0	0	0
3	I- M-	0	0	0
4	rr M+	1+s	2+	2+

**Final interpretation of Panel:**

**Anti-M reactive @ IS, 37 and IAT. Other common alloantibodies ruled out.**



# ABO Discrepancy

<b>Reverse grouping</b>	<b>A<sub>1</sub> cells (M-)</b>	<b>B cells (M-)</b>
<b>Patient J.G.</b>	<b>2+</b>	<b>4+</b>

# ABO Resolution

	Anti-M	Anti-A <sub>1</sub>	Pt. J.G.
Gp A #1	0	4+	2+
Gp A #2	0	0	0
Gp A #3	0	4+	3+

# ABO Resolution

	Anti-M	Anti-A <sub>1</sub>	Pt. J.G.
Gp A #1	0	4+	2+
Gp A #2	0	0	0
Gp A #3	0	4+	3+

**Interpretation: Probable anti-A<sub>1</sub>; unable to find additional M-, A<sub>1</sub>- cells to fully resolve ABO discrepancy**

# J.G. Eluate

	<b>Eluate</b>	<b>Last Wash</b>
<b>SI</b>	<b>0</b>	<b>0</b>
<b>SII</b>	<b>0</b>	<b>0</b>
<b>SIII</b>	<b>0</b>	<b>0</b>
<b>AC (EGA-tr)</b>	<b>1+w</b>	<b>0</b>

# J.G. Eluate

	<b>Eluate</b>	<b>Last Wash</b>
<b>A<sub>1</sub> cells</b>	<b>1+s</b>	<b>0</b>
<b>B cells (x 2)</b>	<b>0</b>	<b>0</b>
<b>A<sub>1</sub> cells</b>	<b>1+s</b>	<b>0</b>
<b>A<sub>2</sub> cells</b>	<b>1+w</b>	<b>0</b>

- Interpretation: Anti-A eluted from patient cells

# Where did that come from?

- No known transfusion of blood products
- Medication history (extensive list):
  - Cefepime
  - Insulin
  - Diphenhydramine injection
  - Multiple antibiotic ointments
  - **IV Immune Globulin**

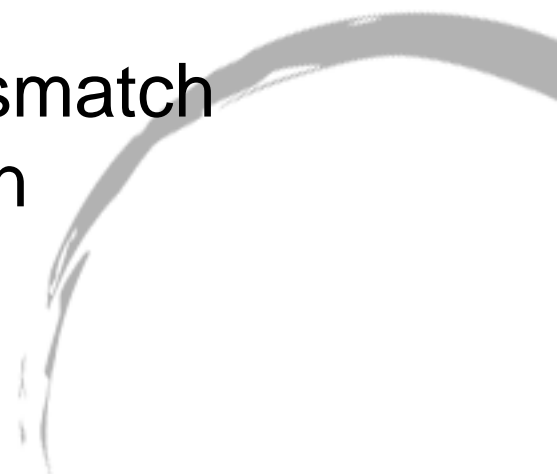
# Antibodies in IVIG

- “Commercial immune globulins, including IVIG, have measurable levels of anti-A and -B (IgG class), as well as a variety of non-ABO antibodies...RBC antibodies have been found in 49% of 165 lots of IVIG, including anti-K, -C and –Le<sup>b</sup>.”
- “..examples have been reported of hemolytic anemias due to anti-D or anti-A.”

Reference: Intravenous immune globulins: an update for clinicians. Transfusion 2003;43:1460-1480



# J.G.

- Summary of results: Anti-A in eluate. Anti-M and apparent anti-A<sub>1</sub> in plasma.
    - Anti-A<sub>1</sub> in plasma may be from the IVIG, and is only reacting with the A<sub>1</sub> cells because the antibody is weak
    - Recommended group O, crossmatch compatible units for transfusion
- 



# Case D.S.

- Patient D.S.
  - Female, race unknown, DOB 11/25/70
  - Dx: Uncontrolled hypertension, anemia
  - Never transfused
  - No medications
  - History of 1 pregnancy

**D.S.**

- Initial results:
  - Group A, Rh positive (no discrepancies)
  - DAT:

<b>PS</b>	<b>-IgG</b>	<b>-C3</b>	<b>Control</b>
<b>2+</b>	<b>Micro+</b>	<b>2+</b>	<b>0</b>

# D.S.

	<b>IS</b>	<b>PEG IAT</b>
<b>SI</b>	<b>0</b>	<b>3+</b>
<b>SII</b>	<b>0</b>	<b>0</b>
<b>SIII</b>	<b>0</b>	<b>1+s</b>
<b>AC</b>	<b>0</b>	<b>3+</b>

# D.S.

- Phenotype

(EGA-treated for AHG typings):

C+E-c+e+; K-; Fy(a+b+); Jk(a+b+);

S+s-; M+N-; Le(a-b-)

# Antibody Screen

	<b>Rh</b>	<b>K</b>	<b>S</b>	<b>s</b>	<b>Le<sup>a</sup></b>	<b>Le<sup>b</sup></b>	<b>M</b>	<b>N</b>
<b>SI</b>	<b>R1R1</b>	<b>+</b>	<b>0</b>	<b>+</b>	<b>0</b>	<b>+</b>	<b>0</b>	<b>+</b>
<b>SII</b>	<b>R2R2</b>	<b>0</b>	<b>+</b>	<b>+</b>	<b>0</b>	<b>+</b>	<b>+</b>	<b>0</b>
<b>SIII</b>	<b>rr</b>	<b>0</b>	<b>+</b>	<b>0</b>	<b>+</b>	<b>0</b>	<b>+</b>	<b>0</b>

# Antibody Screen

	Rh	K	S	s	Le <sup>a</sup>	Le <sup>b</sup>	M	N
SI	R1R1	+	0	+	0	+	0	+
SII	R2R2	0	+	+	0	+	+	0
SIII	rr	0	+	0	+	0	+	0

Rh/ other	K	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	S	s	M	N	Le <sup>a</sup>	Le <sup>b</sup>	PEG IgG
R2R2	0	+	+	0	+	+	0	+	0	0	+	0
rr	+	0	+	0	+	0	+	+	+	0	+	0
rr,Lu(a+)	+	+	+	+	0	+	+	+	0	0	+	2+s
rr	0	+	+	+	+	0	+	0	+	+	0	2+
R1R1	0	0	+	+	+	+	0	+	+	+	0	2+
rr, Lu(a+b-)	0	+	+	+	0	0	+	0	+	0	+	2+s

Rh/ other	K	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	S	s	M	N	Le <sup>a</sup>	Le <sup>b</sup>	PEG IgG
R2R2	0	+	+	0	+	+	0	+	0	0	+	0
rr	+	0	+	0	+	0	+	+	+	0	+	0
rr, Lu(a+)	+	+	+	+	0	+	+	+	0	0	+	2+s
rr	0	+	+	+	+	0	+	0	+	+	0	2+
R1R1	0	0	+	+	+	+	0	+	+	+	0	2+
rr, Lu(a+b-)	0	+	+	+	0	0	+	0	+	0	+	2+s



Rh/ other	K	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	S	s	M	N	Le <sup>a</sup>	Le <sup>b</sup>	PEG IgG
R2R2	0	+	+	0	+	+	0	+	0	0	+	0
rr	+	0	+	0	+	0	+	+	+	0	+	0
rr, Lu(a+)	+	+	+	+	0	+	+	+	0	0	+	2+s
rr	0	+	+	+	+	0	+	0	+	+	0	2+
R1R1	0	0	+	+	+	+	0	+	+	+	0	2+
rr, Lu(a+b-)	0	+	+	+	0	0	+	0	+	0	+	2+s
rr	+	+	+	+	+	0	+	0	+	0	+	2+
R2R2, Lu(a+)	0	0	+	0	+	0	+	+	+	+	0	0

Rh/ other	K	Fy <sup>a</sup>	Fy <sup>b</sup>	Jk <sup>a</sup>	Jk <sup>b</sup>	S	s	M	N	Le <sup>a</sup>	Le <sup>b</sup>	PEG IgG
R2R2	0	+	+	0	+	+	0	+	0	0	+	0
rr	+	0	+	0	+	0	+	+	+	0	+	0
rr,Lu(a+)	+	+	+	+	0	+	+	+	0	0	+	2+s
rr	0	+	+	+	+	0	+	0	+	+	0	2+
R1R1	0	0	+	+	+	+	0	+	+	+	0	2+
rr, Lu(a+b-)	0	+	+	+	0	0	+	0	+	0	+	2+s
rr	+	+	+	+	+	0	+	0	+	0	+	2+
R2R2, Lu(a+)	0	0	+	0	+	0	+	+	+	+	0	0

D.S.

	PEG/IAT
<b>EGA-treated Autologous Cells + plasma</b>	<b>3+</b>
<b>EGA-tr. Cell control (tested w/ 6% alb)</b>	<b>0</b>

- Additional Jk(a-) cells tested to complete rule outs
- Eluate demonstrated Anti-Jk<sup>a</sup>
- **Conclusion: Autoimmune anti-Jk<sup>a</sup> in serum and eluate**

On to more case studies with LeeAnn...



# It's not always what it seems...

- T&S ordered for patient having hysterectomy
- History: 3 pregnancies, transfused after 2<sup>nd</sup> (2 units)
- Initial results
  - O Rh positive, DAT negative

	<b>Gel</b>
<b>SI</b>	<b>1+</b>
<b>SII</b>	<b>1+</b>
<b>SIII</b>	<b>1+</b>
<b>AC</b>	<b>0</b>

	D	C	E	c	e	Cw	K	k	Fya	Fyb	Jka	Jkb	Lea	Leb	P1	M	N	S	s		Gel IgG
1	+	+	0	0	+	+	0	+	0	+	+	+	+	0	+	+	+	+	+		1+
2	+	+	0	0	+	0	+	+	+	+	+	0	0	+	+	+	0	+	0		1+
3	+	0	+	+	0	0	0	+	0	+	+	+	0	+	+	+	+	0	+		1+
4	+	0	0	+	+	0	0	+	0	0	+	+	0	0	+	+	0	+	+		1+
5	0	+	0	0	+	0	0	+	0	+	0	+	0	+	+	+	+	+	+		1+
6	0	0	+	+	+	0	0	+	+	+	+	+	0	+	+	+	0	0	+		1+
7	0	0	0	+	+	0	+	0	0	+	0	+	0	+	0	+	+	0	+		0
8	0	0	0	+	+	0	0	+	+	0	0	+	+	0	+	0	+	+	+		1+
9	0	0	0	+	+	0	0	+	0	+	+	+	0	+	0	+	0	+	0		1+
10	+	+	0	+	+	0	0	+	+	0	+	0	0	+	0	+	+	0	+		1+
11	0	0	0	+	+	0	0	+	+	+	0	+	0	0	+	0	+	+	+		1+
Auto																					0

- What would you do next?

# Selected cells...

	D	C	E	c	e	Cw	K	k	Fya	Fyb	Jka	Jkb	Lea	Leb	P1	M	N	S	s		Gel IgG
1	+	+	0	0	+	+	+	0	+	+	+	+	+	0	+	0	+	+	+		1+
2	+	0	+	+	0	0	+	0	0	+	+	0	0	+	+	+	0	0	+		1+
3	+	0	0	+	+	0	+	0	0	0	+	+	0	0	+	+	0	+	0		1+
4	0	+	0	0	+	0	+	0	0	+	0	+	0	+	+	+	+	0	+		1+

- What now??

- Phenotype

- R<sub>0</sub>r, Kk, Fy(a-b+), Jk(a+b-), Le(a-b+), P1, MNs

	D	C	E	c	e	Cw	K	k	Fya	Fyb	Jka	Jkb	Lea	Leb	P1	M	N	S	s		Gel IgG
1	+	0	0	+	+	0	+	+	+	0	+	0	+	0	+	+	0	0	+		0
2	+	0	0	+	+	0	0	+	+	0	0	+	0	+	0	+	+	0	0		0
3	+	+	0	0	+	0	0	+	+	+	+	0	0	0	+	0	+	0	+		1+
4	+	0	0	+	+	0	0	+	+	+	+	+	0	+	+	+	0	+	0		1+
5	+	0	+	+	+	0	+	+	0	+	0	+	0	+	0	+	+	0	+		1+

- Summary

- Anti-C, -E, -S present
- Anti-Fy<sup>a</sup>, -Jk<sup>a</sup>, -Le<sup>a</sup> ruled out
- No anti-k!
- Let's look at the original panel...





# You really can help us...

- Prenatal patient at XYZ Hospital
  - Dx: placenta previa
- Order for 4 RBCs
- Request form indicates:
  - A, Rh positive
  - IAT positive – 2 of 3 cells

# Reference Lab performs normal workup

	D	C	E	c	e	Cw	K	k	Fya	Fyb	Jka	Jkb	Lea	Leb	P1	M	N	S	s	IS	PEG IgG
1	+	+	0	0	+	+	0	+	0	+	+	+	+	0	+	+	+	+	+	0	0
2	+	+	0	0	+	0	+	+	+	+	+	0	0	+	+	+	0	+	0	0	0
3	+	0	+	+	0	0	0	+	0	+	+	+	0	+	+	+	+	0	+	0	0
4	+	0	0	+	+	0	0	+	0	0	+	+	0	0	+	+	0	+	+	0	0
5	0	+	0	0	+	0	0	+	0	+	0	+	0	+	+	+	+	+	+	0	0
6	0	0	+	+	+	0	0	+	+	+	+	+	0	+	+	+	0	0	+	0	0
7	0	0	0	+	+	0	+	0	0	+	0	+	0	+	0	+	+	0	+	0	0
8	0	0	0	+	+	0	0	+	+	0	0	+	+	0	+	0	+	+	+	0	0
9	0	0	0	+	+	0	0	+	0	+	+	+	0	+	0	+	0	+	0	0	0
10	+	+	0	+	+	0	0	+	+	0	+	0	0	+	0	+	+	0	+	0	0
11	0	0	0	+	+	0	0	+	0	+	0	+	0	0	+	0	+	+	+	0	0
Auto																				0	0

- What should we do next?

# Let's call XYZ hospital!

- Routine technique in XYZ lab is gel!
- What do you think will happen if we test in gel?

# Let's call XYZ hospital!

- Routine technique in XYZ lab is gel!
- What do you think will happen if we test in gel?

	D	C	E	c	e	Cw	K	k	Fya	Fyb	Jka	Jkb	Lea	Leb	P1	M	N	S	s		Gel IgG
1	+	0	0	+	+	0	+	+	+	0	+	0	+	0	+	+	0	0	+		2+
2	+	0	0	+	+	0	0	+	+	0	0	+	0	+	0	+	+	0	0		1+
3	+	+	0	0	+	0	0	+	+	+	+	0	0	0	+	0	+	0	+		0
4	+	0	0	+	+	0	0	+	+	+	+	+	0	+	+	+	0	+	0		2+
5	0	0	0	+	+	0	0	+	0	0	0	+	+	0	0	0	+	+	+		0
6	+	0	+	+	0	0	+	+	0	+	0	+	0	+	0	0	+	0	+		0

# Let's call XYZ hospital!

- Routine technique in XYZ lab is gel!
- What do you think will happen if we test in gel?

	D	C	E	c	e	Cw	K	k	Fya	Fyb	Jka	Jkb	Lea	Leb	P1	M	N	S	s		Gel IgG
1	+	0	0	+	+	0	+	+	+	0	+	0	+	0	+	+	0	0	+		2+
2	+	0	0	+	+	0	0	+	+	0	0	+	0	+	0	+	+	0	0		1+
3	+	+	0	0	+	0	0	+	+	+	+	0	0	0	+	0	+	0	+		0
4	+	0	0	+	+	0	0	+	+	+	+	+	0	+	+	+	0	+	0		2+
5	0	0	0	+	+	0	0	+	0	0	0	+	+	0	0	0	+	+	+		0
6	+	0	+	+	0	0	+	+	0	+	0	+	0	+	0	0	+	+	0		0

# Now what do we know?

- No reactivity in PEG-IgG tests
- Possible anti-M reactive in gel
  - Additional rule outs needed for:
    - Anti-C, -E, -K, -Fy<sup>a</sup>, -Fy<sup>b</sup>, -Jk<sup>a</sup>, -S, -s
- Reactivity of anti-M enhanced in gel
  - Possibly due to slightly lower pH in gel
- Is this significant?
- Could we have arrived at this answer faster with XYZ's testing method?

# Summary

- Working together allows everyone to achieve the best results and provide the best products for our patients.
  - Simple techniques can help with problem solving.
  - Proper sample labeling and completion of request form are critical to sample acceptance.